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NEW AND RARE FUNGI ON ALIEN WOODY PLANTS
IN SVERDLOVSK REGION (THE MIDDLE URALS, RUSSIA)© 2025. O. S. Shiryaeva^{1,*}, T. A. Surina^{2,**}, I. V. Zmitrovich^{3,***}, A. S. Budimirov^{1,****},
T. S. Bulgakov^{4,*****}, L. V. Zaitseva^{2,*****} and A. G. Shiryaev^{1,*****}¹ Institute of Plant and Animal Ecology of the Ural Branch of the Russian Academy of Sciences, 620144 Ekaterinburg, Russia² All-Russian Plant Quarantine Centre, 140150 Bykovo, Russia³ Komarov Botanical Institute of the Russian Academy of Sciences, 197022 St. Petersburg, Russia⁴ Federal Research Centre the Subtropical Scientific Centre of the Russian Academy of Sciences, 354002 Sochi, Russia

*e-mail: olga.s.shiryaeva@gmail.com

**e-mail: t.a.surina@yandex.ru

***e-mail: iv_zmitrovich@mail.ru

****e-mail: budimirov.alex@gmail.com

*****e-mail: ascomycologist@yandex.ru

*****e-mail: lidiya.zaitseva26@yandex.ru

*****e-mail: anton.g.shiryaev@gmail.com

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A groundbreaking study has documented 28 species of phytopathogenic and saprobic fungi, belonging to *Ascomycota* and *Basidiomycota* divisions, on alien trees and shrubs in Sverdlovsk Region for the first time. In addition, 14 rare species — each with only two to four previous sightings in the region — were identified. Notably, four species — *Cheirospora botryospora*, *Homortomyces tamaricis*, *Lophomerum rhododendri*, and *Pyrenopeziza lonicerae* — were recorded in Russia for the first time. The surveyed fungi were found growing on 34 woody plant species, 27 of which occur outdoors while the remaining seven are restricted to greenhouse environments. The study also sheds light on dangerous invasive phytopathogenic fungi in the region, including *Biscogniauxia mediterranea* and *Entoleuca mammata*. Additionally, the invasive species *Hymenoscyphus fraxineus*, recently discovered in Sverdlovsk Region, has been recorded at a new site. The ITS1–5.8S–ITS2 sequence of *H. fraxineus* collected at this location corresponds to the standard East Asian sequence variant. This suggests that local populations of *H. fraxineus* in Sverdlovsk Region may not originate from Europe but rather from East Asia.

Keywords: alien species, *Ascomycota*, *Basidiomycota*, biodiversity, fungal distribution, invasion, plant pathogens

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INTRODUCTION

Fungal diversity in Sverdlovsk Region has been under investigation for over a century, with foundational studies carried out by researchers such as P.V. Syuzev (1898, 1911), N.A. Naumov (1915), Z.A. Demidova (1925, 1970), and N.T. Stepanova-Kartavenko (1967), among others. However, focused research on alien and invasive fungal species in the region has only gained momentum in the last few decades (Shiryaev, 2009; Shiryaev et al., 2010; Shiryaev, Stavishenko, 2011). This study represents the 11th publication in a series dedicated to identifying fungal species presented on alien woody plants in the Middle Urals (Bulgakov, Shiryaev, 2021, 2022; Shiryaev et al., 2021, 2022a, 2022b, 2023a, 2023b, 2023c, 2024; Shiryaev, Kiseleva, 2023). The primary objective is to document newly observed and rare (with one to four previous sightings) species colonizing alien woody plants within the region.

MATERIALS AND METHODS

Materials. The referred species were gathered by A. Shiryaev and T. Bulgakov during the fieldwork in 2015–2025 at different sites within Sverdlovsk Region. In addition, the specimens of phytopathogenic fungi provided in recent years by botanical gardens staff, phytodesigners, and amateurs (N. Azalova, V. Domrachev, P. Martushev, V. Zavorukhin) were studied. This material is stored at the Museum of the Institute of Plant and Animal Ecology (SVER). A number of specimens from the SVER were examined. There are collections of O. Clerc, P. Syuzev, A. Kazanskiy, Z. Demidova, A. Sirko, N. Stepanova, S. Kadochnikov, N. Ushakova, S. Ljashko, B. Panov, and I. Tikhonova, made in the period from the beginning of the 20th century to the present day.

Morphological study. All specimens were examined under light microscope Leica DM 2000. The microscopic features were studied from the material

mounted in distilled water, 10% NH_4OH , 5% KOH, Melzer's reagent and 1% Congo Red in concentrated NH_4OH .

Special key-books and monographs were used for identification of the fungal species (Kuprevich, Ulyanishchev, 1975; Ulyanishchev, 1978; Teterevnikova-Babayan, 1987; Butin, 1989; Braun, Melnik, 1997; Ellis, Ellis, 1997; Braun, 1998; Karatygin, 2002; Braun, Cook, 2012; Knudsen, Vesterholt, 2012; Ryvarden, Melo, 2014; Blagoveshchenskaya, 2015), as well as some additional publications devoted to the study and taxonomical revision of the some particular fungal taxa and new species descriptions (Hyde et al., 2018; Sennawanna et al., 2019; Crous et al., 2020) and open global database "Fungal Databases: U.S. National Fungus Collections" (Farr, Rossman, 2024). Fungal names are given according to the Index Fungorum (2025) online database.

The host plant species were identified by the key-book "Keys to the trees and shrubs species of the Urals" (Mamaev, 2000); the plant species are given according to the open nomenclatural database "Plant of the World Online" (POWO, 2025). Some introduced exotic plant species in the collections of the Botanical garden were determined according to the personal data of researchers of the Botanical Garden of the Ural Branch of the Russian Academy of Sciences.

The following abbreviations are used in the list: new species for Russia are marked with an exclamation mark; new species for Sverdlovsk Region are marked with an asterisk; Ekb – Ekaterinburg City; BCD – Botanical City District; BG UrB RAS – Botanical Garden of the Ural Branch of the Russian Academy of Sciences; BG UFU – Botanical Garden of the Ural Federal University; DP – Dendrarium at Pervomayskaya street; D8 – Dendrarium at 8 March street; MCP – Maykovskiy Central Park of Ekaterinburg city; VG – Prof. L.I. Vigorov Medicinal crops garden.

DNA extraction, amplification, and sequencing. One apothecium from the specimen tentatively assigned to *Hymenoscyphus fraxineus* was used for the DNA extraction. DNA extraction was performed using the MagnoPrime PHYTO reagent kit (NextBIO) according to the manufacturer's instructions.

The ITS1–5.8S–ITS2 region of rDNA was amplified with the universal primer pair ITS5/ITS4 (White et al., 1990). PCR reaction was carried out under the following conditions: initial denaturation for 3 min at 95°C, 40 cycles (denaturation for 30 s at 95°C, primers annealing for 30 s at 52°C, extend DNA for 30 s at 72°C), final extension for 7 min at 72°C. PCR products intended for sequencing were purified using the commercial GeneJET Genomic DNA Purification Kit (Thermo Scientific). The sequencing reaction was performed using BigDye Terminator v. 3.1 Cycle Sequencing Kit reagents (Applied Biosystems) according to the manufacturer's instructions, followed by

fragment separation on an Applied Biosystems 3500 Genetic Analyzer. Sequencing of the PCR products was performed with the same primer pairs used for PCR amplification.

Newly generated sequence was deposited in GenBank (PX251630). Molecular studies were performed at the All-Russian Plant Quarantine Center (VNIICR).

Phylogenetic analysis. Thirty-six fungal ITS sequences were retrieved from GenBank to determine the phylogenetic placement of *Hymenoscyphus* specimens (Table 1). *H. fructigenus* was chosen as the out-group (Gross, Han, 2015). Sequence alignments were constructed in MEGA X (Kumar et al., 2018) using MUSCLE algorithm (Edgar, 2004) and trimmed manually. Phylogenetic analysis was conducted using Bayesian Inference (BI) and Maximum Likelihood (ML). ML were performed with RAXML GUI v. 2.0.14. The nodal support for the individual branches was estimated by bootstrapping using 1000 replicates. Bayesian analyses were conducted with BEAST v. 2.7.6 using Yule tree prior and strict molecular clocks (Bouckaert et al., 2014). A single chain of 10^7 steps was run with a burn-in of 25%.

RESULTS

An annotated species list

Ascomycota

Dothideomycetes

Botryosphaerales

Botryosphaeriaceae

**Diplodia lonicerae* Fuckel – Ekb, MCP, on *Lonicera tatarica* L., coll. N. Azalova, det. A. Budimirov, T. Bulgakov, 09.07.2020, SVER(F) 17325.

Dothiorella sarmentorum (Fr.) A.J.L. Phillips, A. Alves et J. Luque (= *Diplodia melaena* Lév.; = *D. pruni* Fuckel) – Kamensk area, artificial treeline along the road "Kamensk-Uralsk-Ekaterinburg", on *Ulmus pumila* L., coll. N. Ushakova, det. A. Budimirov, T. Bulgakov, 21.09.1999, SVER(F) 17363; Ekb, suburb cottage village Palnix, on *Prunus armeniaca* L., coll. V. Zavorukhin, det. A. Budimirov, 25.05.2023, SVER(F) 17338. There are two new records in the region. The species was mentioned once before (Shiryaev et al., 2024).

Mycosphaerellales

Mycosphaerellaceae

Neophloeospora maculans (Bérenger) Videira et Crous [= *Mycosphaerella mori* (Fuckel) F.A. Wolf; = *Septoria mori* Fuckel] – Ekb, suburb cottage village Aleksandriya, on leaves of *Morus alba* L., coll. N. Azalova, det. A. Budimirov, T. Bulgakov, 26.09.2021, SVER(F) 17364. This is the second record in the region, the species has been mentioned earlier (Shiryaev et al., 2023c).

**Ruptoseptoria unedonis* (Roberge ex Desm.) Quaedvl., Verkley et Crous (= *Septoria unedonis* Roberge ex Desm.) – Ekb, greenhouse at the M.A. Nurov estate, on leaves of *Arbutus andrachne* L., coll. A. Kazanskiy, Z. Demidova, det. A. Budimirov, T. Bulgakov, 19.07.1928, SVER(F) 17337.

**Septoria aesculina* Thüm. – Ekb, BG UrB RAS, on leaves of *Aesculus hippocastanum* L., coll. N. Azalova, det. A. Budimirov, T. Bulgakov, 30.09.2020, SVER(F) 17339.

S. elaeagni (Chevall.) Desm. (= *Depazea elaeagni* Chevall.) – Beloyarsk area, artificial treeline along the road, on leaves of *Elaeagnus angustifolia* L., coll. A. Sirko, det. A. Budimirov, T. Bulgakov, 04.10.1962, SVER(F) 17361. This is the second record in the region, the species has been mentioned earlier (Shiryaev, 2024).

Table 1. Voucher information and GenBank accession numbers for ITS sequences used in this study

Таблица 1. Информация о ваучерных образцах и регистрационных номерах GenBank для последовательностей ITS, использованных в данном исследовании

Species (current name)	Specimen/Isolate name	ITS	Location	Sequence origin
<i>Hymenoscyphus fraxineus</i> (T. Kowalski) Baral, Queloz et Hosoya	SVER(F) 17334	PX251630	Russia, the Middle Urals	this study
<i>H. fraxineus</i>	8096	KF188724	China	Drenkhan et al. (2017)
“ “	EST3146-2	KR733591	Russia, Far East	“ “
“ “	EST3144	KP403806	Russia, Far East	“ “
“ “	EST2723	KR733600	Russia, Far East	“ “
“ “	EST3145	KP403807	Russia, Far East	“ “
“ “	EST3146	KR733592	Russia, Far East	“ “
“ “	080517.2	FJ429378	Lithuania	Ioos et al. (2009)
“ “	08-410	FJ429376	France	“ “
“ “	080520	FJ429379	Lithuania	“ “
“ “	1582	FJ429380	Poland	“ “
“ “	CBS 122504	FJ597975	Poland	Kowalski, Holdenrieder (2009)
“ “	KUS-F52255_1	KP068053	South Korea	Schoebel et al. (2014)
“ “	PRM:857180	HF937557	Slovakia	Koukol et al. (2016)
“ “	KUS-F52613	KF830853	South Korea	Han et al. (2014)
“ “	TNS:F-40043	AB926104	Japan	unpublished
<i>H. albidus</i> (Gillet) W. Phillips	CBS 126533	MH864151	Switzerland	Böhm et al. (2024)
“ “	EST2591	KP403803	Lithuania	“ “
“ “	CNRHalbLDPRO	MW238522	Italy	“ “
“ “	LOU-M-8	KC481686	France	“ “
<i>H. linearis</i> Hosoya, Andr. Gross et Baral	Chic_01	KM114521	Japan	Gross et al. (2015)
“ “	Chic_03	KM114523	Japan	“ “
<i>H. koreanus</i> Andr. Gross et J.G. Han	KUS F52847_01	KP068057	South Korea	“ “
“ “	KUS-F52742_2	KP068074	South Korea	“ “
“ “	KUS-F52542_2	KP068078	South Korea	“ “
<i>H. albidoides</i> H.D. Zheng et W.Y. Zhuang	HMAS 264141	KF188721	China	Zheng, Zhuang (2014)
“ “	HMAS 264140	KF188722	China	“ “
“ “	HMAS 264142	KF188723	China	“ “
<i>H. pusillus</i> T. Kowalski et Bilański	HMC 21525	MH476516	Poland	Kowalski, Bilański (2019)
“ “	HMC 21521	MH476512	Poland	“ “
“ “	HMC 21523	MH476514	Poland	“ “
“ “	HMC 21527	MH476518	Poland	“ “
<i>H. occultus</i> Andr. Gross et J.G. Han	KUS-F52847	KP068058	South Korea	Gross, Han (2015)
“ “	KUS-F52847_22	KP068062	South Korea	“ “
“ “	KUS-F52847_24	KP068063	South Korea	“ “
“ “	KUS-F52847_25	KP068064	South Korea	“ “
<i>H. fructigenus</i> (Bull.) Gray	CBS 650.92	GU586933	Germany	“ “

**S. oleandrina* Sacc. — Ekb, greenhouse at the M.A. Nurov estate, on leaves of *Nerium oleander* L., coll. A. Kazanskiy, Z. Demidova, det. T. Bulgakov, A. Budimirov, 10.06.1928, SVER(F) 17365.

**S. robiniae* (Lib.) Desm. [= *Phloeospora robiniae* (Lib.) Höhn.] — Ekb, VG, on leaves of *Robinia pseudoacacia* L., coll. A. Shiryaev, det. T. Bulgakov, A. Budimirov, 18.09.2023, SVER(F) 17362.

Sphaerulina neoaceris Crous et Bulgakov — Ekb, Elizavet's church, on *Acer negundo*, coll. O. Clerc, det. A. Budimirov, T. Bulgakov, 01.09.1916, SVER(F) 17360. This is the earliest record of the species in the region. A later record has been already published in a previous paper (Bulgakov, Shiryaev, 2021).

**S. menispermi* (Thüm.) Quaedvl., Verkley et Crous (= *Septoria menispermi* Thüm.) — Ekb, VG, on leaves of *Menispermum dahuricum* DC., coll. N. Stepanova, det. A. Budimirov, T. Bulgakov, 24.08.1980, SVER(F) 17336.

Wilsonomyces carpophilus (Lév.) Adask., J.M. Ogawa et E.E. Butler [= *Stigmata carpophila* (Lév.) M.B. Ellis; = *Thyrostroma carpophilum* (Lév.) B. Sutton] — Ekb, greenhouse of the Turbomotor Plant, on leaves of *Prunus laurocerasus*, coll. S. Kadochnikov, det. A. Budimirov, T. Bulgakov, 06.06.1994, SVER(F) 17340; *ibid.*, suburb cottage village Palnix, on *Prunus armeniaca* L., coll. N. Azalova, det. A. Budimirov, T. Bulgakov, 25.05.2023, SVER(F) 17359. The species was previously known in the region from two records (Bulgakov, Shiryayev, 2021).

Thyridariaceae

Thyridaria macrostomoides (De Not.) M.E. Barr [= *Lophiostoma macrostomoides* (De Not.) De Not.] — Ekb, MCP, on *Populus balsamifera* L., coll. A. Shiryayev, det. A. Budimirov, T. Bulgakov, 02.08.2021, SVER(F) 17358. The species was previously recorded on aspens and black poplars within the natural habitats of the region (Stepanova, Sirko, 1970).

Homortomycetales

Homortomycetaceae

!**Homortomyces* cf. *tamaricis* Wijayaw., Camporesi et K.D. Hyde — Ekb, BG UrB RAS, on *Tamarix ramosissima* Ledeb., coll. A. Shiryayev, det. T. Bulgakov, A. Budimirov, 30.05.2021, SVER(F) 17326.

Pleosporales

Camarosporidiellaceae

Camarosporidiella neomori Jaklitsch et Voglmayr [= *C. moricola* (Chethana, Bulgakov et K.D. Hyde) Wanas. et K.D. Hyde] — Ekb, suburb cottage village Palnix, on *Morus alba*, coll. N. Azalova, det. T. Bulgakov, 14.09.2023, SVER(F) 17335. This is the second record in the region, the species has been mentioned earlier (Shiryayev, 2024).

Didymellaceae

Neodidymelliopsis negundinis Manawas., Camporesi et K.D. Hyde — Krasnoufmsk town, terrace along the Ufa river, on *Acer negundo* L., coll. A. Shiryayev, det. T. Bulgakov, 03.09.2020, SVER(F) 17341. This is the third record in the region. The species was previously known from two records (Bulgakov, Shiryayev, 2022).

Leptosphaeriaceae

Longiseptatispora curvata Crous et Bulgakov [= *Rhabdospora lonicerae* (Cooke et Ellis) Sacc.] — Krasnoufmsk area, surrounds of Natalyinsk village, artificial treeline along the road, on dying twigs of *Lonicera tatarica* L., coll. A. Shiryayev, det. T. Bulgakov, 02.09.2023, SVER(F) 17357. This is the third record in the region. The species was previously known from two records (Shiryayev et al., 2023c).

Leotiomycetales

Helotiales

Erysiphaceae

**Arthrocladiella mougeotii* (Lév.) Vassilkov — Ekb, glasshouse of the Turbomotor Plant, on leaves of *Lycium barbarum* L., coll. S. Ljashko, det. A. Budimirov, 09.06.1993, SVER(F) 17342.

**Podosphaera leucotricha* (Ellis et Everh.) E.S. Salmon — Ekb, greenhouse at the M.A. Nurov estate, on leaves of *Pyrus malus* L., coll. O. Clerc, det. A. Budimirov, 26.05.1914, SVER(F) 17327.

Helotiaceae

Hymenoscyphus fraxineus (T. Kowalski) Baral, Queloz et Hosoya — Kamenskiy area, artificial treeline along the road, on fallen petioles of *Fraxinus excelsior* L., coll. V. Domrachev, det. T. Surina et L. Zaitseva, 02.08.2024, SVER(F) 17334. This is the third record in the region, the species has been mentioned earlier for Ekaterinburg city (Shiryayev, Kiseleva, 2023).

Note. *H. fraxineus* group now consists of seven species (Kowalski, Bilański, 2019). They are morphologically similar but clearly separated in phylogeny. Identification of specimen was confirmed using molecular methods. The specimen was grouped with other *H. fraxineus* strains with strong nodal support (Fig. 1).

Drepanopezizaceae

**Theadonia ligustrina* (Boerema) B. Sutton — Ekb, suburb cottage village Karasyeozerskiy-2, on *Ligustrum vulgare* L., coll. S. Lyashko, det. T. Bulgakov, A. Budimirov, 21.09.2002, SVER(F) 17333.

Ploettnerulaceae

!**Pyrenopeziza lonicerae* Nannf. — Ekb, BG UFU, on *Lonicera chrysantha* Turcz. ex Ledeb., coll. P. Martushev, det. T. Bulgakov, A. Budimirov, 09.09.2024, SVER(F) 17356.

Incertae sedis

**Monostichella robergei* (Desm.) Höhn. — Ekb, suburb cottage village Karasyeozerskiy-2, on *Carpinus betulus* L., coll. V. Zavorukhin, det. T. Bulgakov, A. Budimirov, 22.09.2024, SVER(F) 17343.

Rhytismatales

Rhytismataceae

!**Lophomerum rhododendri* (Ces. ex Rehm) Remler — Ekb, BG UrB RAS, on *Rhododendron caucasicum* L., coll. A. Shiryayev, det. T. Bulgakov, A. Budimirov, 27.09.2022, SVER(F) 17355.

Sordariomycetes

Diaporthales

Gnomoniaceae

**Apiognomonina hystrix* (Tode) Sogonov [= *Gnomonia cerastis* (Riess) Ces. et De Not.] — Ekb, BCD, on dead fallen leaves of *Acer negundo* L., coll. A. Shiryayev, det. T. Bulgakov, 16.05.2025, SVER(F) 17332.

Melanconidaceae

**Melanconium populinum* Peck — Krasnoufmsk area, artificial treeline along the road, on *Populus balsamifera*, coll. A. Shiryayev, det. T. Bulgakov, 05.09.2023, SVER(F) 17354; Ekb, BCD, on *P. balsamifera*, coll. et det. T. Bulgakov, 13.09.2024, SVER(F) 17344; *ibid.*, Southwestern forest city park, on *P. balsamifera*, coll. et det. T. Bulgakov, 27.07.2022, SVER(F) 17366.

Xylariales

Graphostromataceae

**Biscogniauxia* cf. *dennisii* (Pouzar) Pouzar — Ekb, DP, on branches of *Quercus mongolica* Fisch. ex Ledeb., coll. N. Azalova, det. A. Budimirov, T. Bulgakov, 26.06.2021, SVER(F) 17353.

**B. mediterranea* (De Not.) Kuntze — Ekb, DP, on branch of *Juglans regia* L., coll. B. Panov, det. T. Bulgakov, A. Budimirov, 17.09.2000, SVER(F) 17345.

**B. mandshurica* Lar.N. Vassiljeva — Ekb, BG UrB RAS, on branch of *Malus mandshurica* (Maxim.) Kom., coll. N. Azalova, det. A. Budimirov, T. Bulgakov, 02.10.2022, SVER(F) 17331.

Hypoxylaceae

**Daldinia childiae* J.D. Rogers et Y.M. Ju — Ekb, MCP, on branch of *Fraxinus pensylvanica* Marshall, coll. A. Shiryayev, det. A. Budimirov, T. Bulgakov, 17.09.2023, SVER(F) 17352.

Xylariaceae

**Entoleuca mammata* (Wahlenb.) J.D. Rogers et Y.M. Ju — Ekb, Shartash forest park, on branches of *Populus suaveolens* Fisch., coll. V. Zavorukhin, det. T. Bulgakov, A. Budimirov, 03.08.2021, SVER(F) 17346.

Incertae sedis

!**Cheirospora botryospora* (Mont.) Berk. et Broome [= *Thyrsidium hedericola* (De Not.) Durieu et Mont.] — Ekb, greenhouse at the I.I. Simanov estate, on *Hedera helix* L., coll. O. Clerc, det. N. Naumov, 29.05.1897, SVER(F) 17328.

Basidiomycota

Agaricomycetes

Agaricales

Mycenaceae

**Mycena pseudocorticola* Kühner — Ekb, BG UrB RAS, on bark of *Quercus robur* L., coll. A. Shiryayev, det. O. Shiryayeva, 03.09.2023, SVER 910264.

Pleurotaceae

**Hohenbuehelia cyphelliformis* (Berk.) O.K. Mill. — Ekb, BG UrB RAS, on decayed stump of *Parthenocissus quinquefolia* (L.) Planch., coll. A. Shiryayev, det. O. Shiryayeva, 18.07.2015, SVER 910267.

Resupinataceae

**Resupinatus trichotis* (Pers.) Singer — Ekb, BG UrB RAS, on decayed stump of *Amelanchier* sp., coll. A. Shiryayev, det. O. Shiryayeva, 10.09.2023, SVER 910265.

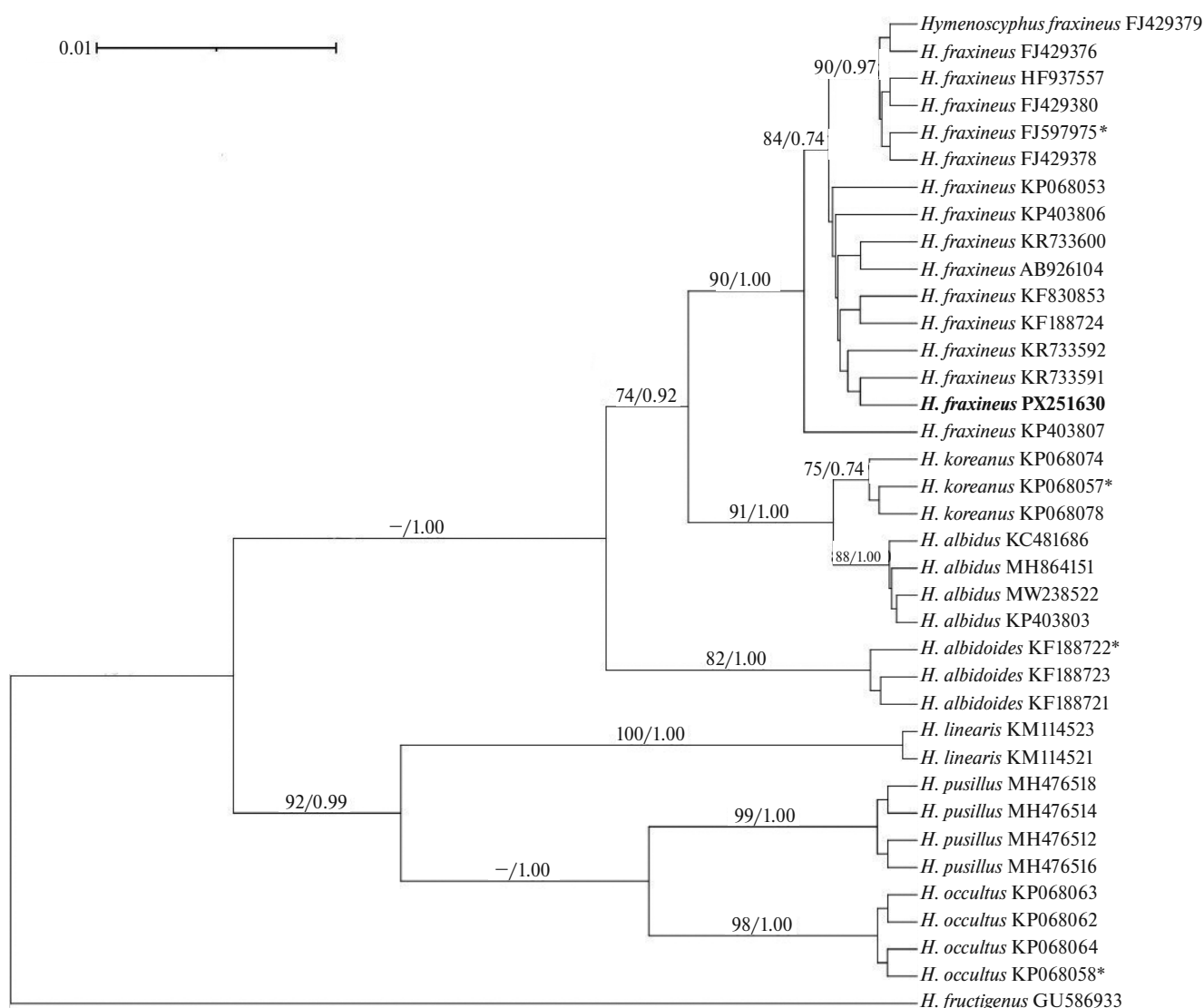


Fig. 1. Phylogram obtained from Bayesian analyses of ITS rDNA data for selected species in the *Hymenoscyphus fraxineus* group. Bootstrap percentages of Maximum Likelihood (with values $\geq 70\%$) are given above the branches in the left, while posterior probabilities of Bayesian analysis are given above the branches in the right. Sequence obtained during this study is presented in bold type. The asterisk indicates the type strains.

Рис. 1. Филограмма, полученная в результате байесовского анализа данных ITS рДНК для группы *Hymenoscyphus fraxineus*. Бутстреп-поддержка (ML) со значениями $\geq 70\%$ приведена над ветвями слева, апостериорная вероятность (BI) – над ветвями справа. Последовательность, полученная в ходе данного исследования, выделена жирным шрифтом. Звездочкой обозначены типовые штаммы.

Polyporales

Fomitopsidaceae

Antrodia kuziana (Pilát) Spirin et Vlasák [= *Brunneoporus malicola* (Berk. et M.A. Curtis) Audet] – Ekb, BG UrB RAS, on the trunk of *Malus baccata* L., coll. et det. A. Shiryayev, 09.09.2024, SVER(F) 17351. This is the fourth record in the region; the previous records were published earlier (Shiryayev et al., 2010).

**Cyanosporus luteocaesius* (A. David) B.K. Cui, L.L. Shen et Y.C. Dai – Ekb, BG UrB RAS, on dead branch of *Pinus mugo* Turra, coll. A. Shiryayev, det. I. Zmitrovich, 02.10.2023, SVER(F) 17347.

Phanerochaetaceae

**Hyphodermella rosae* (Bres.) Nakasone – Ekb, greenhouse at the Rastorguev-Kharitonov estate, on the roots and the base of *Buxus colchica* Pojark., coll. I. Tikhonova, det. A. Shiryayev, 02.07.1990, SVER(F) 17350.

Polyporaceae

**Ganoderma valesiacum* Boud. – Ekb, BG UrB RAS, on the base of alive *Larix kurlensis* Mayr, coll. et det. A. Shiryayev, 03.06.2020, SVER(F) 17348.

Neofavolus suavissimus (Fr.) Seelan, Justo et Hibbett – Ekb, Obroshenskoe Forestry, on decayed stump of *Quercus robur* L., coll. A. Shiryayev, det. O. Shiryayeva, 23.07.2023, SVER 910266. This is the second record in the region, but the decaying wood of *Quercus* is an unusual substrate for this species. *Neofavolus suavissimus* typically decomposes wood of aspens, poplars, and willows. The species has previously been recorded on aspens within the natural habitats of the region (Mukhin et al., 2003).

Steccherinaceae

**Flaviporus brownii* (Humb.) Donk – Ekb, greenhouse at the M.A. Nurov estate, “on the wall of a tub with a palm from Batumi”, coll. P. Syuzev, det. N. Naumov, 06.07.1894, SVER(F) 17329.

Loweomyces wynneae (Berk. et Broome) Jülich – Ekb, at the base of broken trunk of *Juglans mandshurica* Maxim., coll. A. Shiryayev, det. I. Zmitrovich, 17.09.2023, SVER(F) 17349. This is the third record in the region; the species has been mentioned earlier (Shiryayev et al., 2010).

Pucciniomycetes

Pucciniales

Pucciniaceae

Puccinia mirabilissima Peck – Ekb, D8, on leaves of *Mahonia aquifolium* (Pursh) Nutt. (= *Berberis aquifolium* Pursh), 03.09.2018, coll. N. Azalova, det. T. Bulgakov, SVER(F) 17330. This is third record in the region, the species has been mentioned earlier (Shiryaev et al., 2023).

DISCUSSION

A total of 42 species of phytopathogenic and saprobic fungi, belonging to the *Ascomycota* and *Basidiomycota* divisions, were identified growing on alien trees and shrubs. Among these, 28 species are newly recorded in Sverdlovsk Region, while 14 are considered rare, with only two to four documented occurrences in the area. Additionally, four species – *Cheirospora botryospora*, *Homortomyces tamaricis*, *Lophomerium rhododendri*, and *Pyrenopeziza lonicerae* – are being reported in Russia for the first time.

Fungi were found on 34 species of alien woody plants, including 27 species growing in open ground (*Acer negundo*, *Aesculus hippocastanum*, *Carpinus betulus*, *Elaeagnus angustifolia*, *Fraxinus excelsior*, *F. pennsylvanica*, *Juglans mandshurica*, *J. regia*, *Larix kurilenensis*, *Ligustrum vulgare*, *Lonicera chrysantha*, *L. tatarica*, *Mahonia aquifolium*, *Malus baccata*, *M. mandshurica*, *Menispermum dahuricum*, *Morus alba*, *Pinus mugo*, *Populus balsamifera*, *Populus suaveolens*, *Prunus armeniaca*, *Quercus mongolica*, *Q. robur*, *Rhododendron caucasicum*, *Robinia pseudoacacia*, *Tamarix ramosissima*, *Ulmus pumila*), as well as seven species growing in greenhouses (*Arbutus andrachne*, *Buxus colchica*, *Hedera helix*, *Lycium barbarum*, *Nerium oleander*, *Prunus laurocerasus*, *Pyrus malus*). Fungi were collected from different parts of the trees and shrubs, such as leaves, twigs, bark, and wood.

Three fungal species (*Hymenoscyphus fraxineus*, *Entoleuca mammata*, and *Biscogniauxia mediterranea*) are the dangerous invasive pathogens of woody plants (Beenken, Senn-Irlet, 2016; Rabitsch, Nehring, 2021; Voglmayr et al., 2023).

Hymenoscyphus fraxineus is known as an invasive species causing the ash dieback disease (*Fraxinus* spp.), also known as Chalara ash dieback, which was named after *Chalara fraxinea* T. Kovalski – the widely used name of the anamorphic stage of *Hymenoscyphus fraxineus* (Pacia et al., 2023; Zvyagintsev et al., 2023). This harmful phytopathogenic fungus was proposed to be accidentally introduced from East Asia (where it affects native *Fraxinus mandshurica* Rupr.) to Europe in the early 21st century (Zhao et al., 2013) and quickly spread throughout the range of *F. excelsior* and other ash species in most European countries, causing massive dying of ash trees in natural and artificial forests (Pacia et al., 2023). During the 2000s and 2010s, *Hymenoscyphus fraxineus* expanded its range eastward and reached Moscow Region in 2014 (Musolin et al., 2017). After the long-term fixed-route survey, ash dieback

disease caused by *H. fraxineus* was found on the territory of 31 regions in Russia now (Zvyagintsev et al., 2023). Thus, ash dieback now appeared to be a widespread pathology of *Fraxinus* species in their range in most regions of European Russia, including the North Caucasus and Middle Volga. Recent studies suggest that *H. fraxineus* may not be a virulent shoot pathogen on Asian ash species in their native range; moreover, that the current ash dieback epidemic in Europe may not originate directly from the East Asia, or European strains of *H. fraxineus* represent rare genetic variants that have not yet been detected in East Asia (Drenkhan et al., 2017).

The ITS1–5.8S–ITS2 sequence obtained from material collected during the present study differs from the holotype strain of *H. fraxineus* originating from Poland (FJ597975) and from the other European materials by an insertion at position 83 (G instead of a gap) and a point mutation at position 124 (C instead of T) within the ITS1 region. All sequences of *H. fraxineus* originating from the Russian Far East, China, Korea, and Japan were conserved at these two sites (Drenkhan et al., 2017). So, the local *H. fraxineus* strain has conspicuous differences in ITS rDNA sequences from European populations of this species and shows the standard East Asian sequence variant. This suggests that local populations of *H. fraxineus* in Sverdlovsk Region may not originate from Europe but rather from East Asia. Further research is needed to investigate the genetic diversity populations of *H. fraxineus* associated with ash species in the Urals and the virulence of the species.

Entoleuca mammata [= *Hypoxyylon pruinaum* (Klotzsch) Cooke] is a well-known invasive fungal plant pathogen, accidentally introduced from North America to Europe until the 20th century (probably in the 18th century) (Kasanen et al., 2004). This fungus causes the poplar disease known as Hypoxylon cancer, affecting mainly species of the genus *Populus* section *Populus*, most often aspens (Ostry, 2013). This pathogen of poplars is widespread now in most European countries but is especially common in Northern and Western Europe, where it forms a specific fungus-plant pathosystem (Ostry, Anderson, 2009). In Russia, *Entoleuca mammata* affects common aspen (*Populus tremula* L.) and white poplar (*P. alba* L.), causing slow dying of middle-aged and older trees in natural forests throughout the range of aspen and white poplar in European Russia (Bulgakov et al., 2014), the Urals, and Siberia (Kuzmichev et al., 2001). So, the records of this species in Sverdlovsk Region are quite expected.

Biscogniauxia mediterranea is a widespread endophytic fungus that causes cancer and wood rot (Charcoal disease) of many woody plants (Ranjana, Basu, 2012). This species is most common in subtropical Mediterranean countries from Portugal to Iran (Ju et al., 1998; Costa et al., 2022; Kazemzadeh et al., 2021), but is also recorded in East Asia (Ju et al., 1998), South Africa (van der Merwe et al., 2021), and North America (Spaulding, 1961). According to numerous studies,

B. mediterranea can affect many woody plants (Ju et al., 1998; Ragazzi et al., 2012; Ranjana, Basu, 2012), but most often it infects Fagales species (*Fagus*, *Quercus*, *Juglans* and other tree species). The records of this species on walnut (*Juglans regia*) in Sverdlovsk Region can be considered a result of accidental introduction, as well as global warming and aridization of the regional climate, which favor to the invading of subtropical fungal species into the boreonemoral lands of the Urals and further naturalization of alien fungi.

Thus, the present paper supplements information on the phytopathogenic and saprobic fungi (*Ascomycota*, *Basidiomycota*) growing on alien trees and shrubs, including dangerous invasive species.

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REFERENCES

- Beenken L., Senn-Irlet B. Neomyceten in der Schweiz. Stand des Wissens und Abschätzung des Schadpotentials der mit Pflanzen assoziierten gebietsfremden Pilze. WSL Berichte 50. Birmensdorf: Eidg. Forschungsanstalt für Wald, Schnee und Landschaft WSL. 2016.
- Blagoveshchenskaya E. Yu. Phytopathogenic micromycetes: educational key-book. Moscow, 2015. (In Russ.).
- Böhm J.W., Zübert C., Kahlenberg G. et al. Load of the ash dieback pathogen *Hymenoscyphus fraxineus* differs in soil. J. Plant Diseases Protect. 2024. V. 131. P. 1399–1410. <https://doi.org/10.1007/s41348-024-00970-6>
- Bouckaert R., Heled J., Kühnert D. et al. BEAST 2: A Software Platform for Bayesian Evolutionary Analysis. PLOS Computational Biol. 2014. V. 10 (4). Art. e1003537. <https://doi.org/10.1371/journal.pcbi.1003537>
- Braun U. A monograph of *Cercospora*, *Ramularia* and allied genera (phytopathogenic *Hyphomycetes*). V. 2. IHW-Verlag, Eching bei München, 1998.
- Braun U., Cook R.T.A. Taxonomic manual of the *Erysiphales* (powdery mildews). CBS Biodiversity series. V. 11. APS Press, Utrecht, 2012.
- Braun U., Melnik V.A. Cercosporoid fungi from Russia and adjacent countries. Proceedings of the Komarov Botanical Institute. V. 20. Russian Academy of Sciences, SPb., 1997.
- Bulgakov T.S., Shiryayev A.G. New finds of phyllostrophic plant pathogenic microfungi in Ekaterinburg city and its suburbs. Mikologia i fitopatologiya. 2021. V. 55 (6). P. 405–410. <https://doi.org/10.31857/S0026364821060064>
- Bulgakov T.S., Shiryayev A.G. Powdery mildews (*Erysiphaceae*) on woody plants in urban habitats of Sverdlovsk Region (Russia). Mikologia i fitopatologiya. 2022. V. 56 (5). P. 323–331. <https://doi.org/10.31857/S002636482205004x>
- Bulgakov T.S., Vasilyev N.P., Zmitrovich I.V. Summarizing of 10-years investigation on mycobiota of alien trees and shrubs in arboretum of the Otradnoye Research Station of the Komarov Botanical Institute // Botany: history, theory, practice (on the 300th anniversary of the founding of the Komarov Botanical Institute of the Russian Academy of Sciences): Proceedings of the international scientific conference / Ed. D. V. Geltman. LETI, SPb., 2014, pp. 31–39. (In Russ.).
- Butin H. Krankheiten der Wald- und Parkbaume: Diagnose, Biologie, Bekämpfung. Stuttgart etc., 1989.
- Costa D., Ramos V., Tavares R.M. et al. Phylogenetic analysis and genetic diversity of the xylariaceous ascomycete *Biscogniauxia mediterranea* from cork oak forests in different bioclimates. Scientific Reports. 2022. V. 12 (1). P. 2646. <https://doi.org/10.1038/s41598-022-06303-7>
- Crous P.W., Wingfield M.J., Schumacher R.K. et al. New and Interesting Fungi. 3. Fungal Systematics and Evolution. 2020. V. 6. P. 157–231. <https://doi.org/10.3114/fuse.2020.06.09>
- Demidova Z.A. On smut fungi of the Urals. Proc. IPAE. 1970. V. 70. P. 53–60. (In Russ.).
- Demidova Z.A. Overview of diseases of cultivated and wild plants in Ural Oblast. Bull. Uraloblz. 1925. V. 10. P. 9–20. (In Russ.).
- Drenkhan R., Solheim H., Bogacheva A. et al. *Hymenoscyphus fraxineus* is a leaf pathogen of local *Fraxinus* species in the Russian Far East. Plant Pathol. 2017. V. 66. P. 490–500.
- Edgar R.C. MUSCLE: multiple sequence alignment with high accuracy and high throughput. Nucleic Acid Res. 2004. V. 32(5). P. 1792–1797. <https://doi.org/10.1093/nar/gkh340>
- Ellis M.B., Ellis J.P. Microfungi on land plants: an identification handbook. New enlarged edition. Richmond P.C., Slough, 1997.
- Farr D.F., Rossman A.Y. Fungal Databases, U.S. National Fungus Collections, ARS, USDA. 2024.
- Gross A., Han J.G. *Hymenoscyphus fraxineus* and two new *Hymenoscyphus* species identified in Korea. Mycol. Progress. 2015. V. 14. Art. 19. <https://doi.org/10.1007/s11557-015-1035-1>
- Gross A., Hosoya T., Zhao Y.J. et al. Baral *Hymenoscyphus linearis* sp. nov: another close relative of the ash dieback pathogen *H. fraxineus*. Mycol. Progress. 2015. V. 14. Art. 20. <https://doi.org/10.1007/s11557-015-1041>
- Han J.G., Shrestha B., Hosoya T. et al. First report of the ash dieback pathogen *Hymenoscyphus fraxineus* in Korea. Mycobiology. 2014. V. 42 (4). P. 391–396.
- Hyde K.D., Chaiwan N., Norphanphoun C. et al. Mycosphere notes 169–224. Mycosphere. 2018. V. 9 (2). P. 271–430. <https://doi.org/10.5943/mycosphere/9/2/8>
- Index Fungorum. CABI Bioscience, 2025. <http://www.indexfungorum.org>. Accessed 25.02.2025.
- Ioos R., Kowalski T., Husson C. et al. Rapid in planta detection of *Chalara fraxinea* by a real-time PCR assay using a dual-labelled probe. Eur. J. Plant Pathol. 2009. V. 125. P. 329–335.
- Ju Y.-M., Rogers J.D., San Martin F. et al. The genus *Biscogniauxia*. Mycotaxon. 1998. V. 66. P. 1–98.

- <https://doi.org/10.5555/19981005332>
- Karatygin I.V. Key-book to fungi of Russia. The orders *Taphrinales*, *Protomycetales*, *Exobasidiales*, *Microstromatales*. Nauka, SPb., 2002. (In Russ.).
- Kasanen R., Hantula J., Ostry M. et al. North American populations of *Entoleuca mammata* are genetically more variable than populations in Europe. *Mycol. Res.* 2004. V. 108 (7). P. 766–774.
<https://doi.org/10.1017/s0953756204000334>
- Kazemzadeh C.M., Mohammadi H., Sohrabi M. Occurrence and pathogenicity of *Jattaea algeriensis* (*Calosphaeriales*, *Calosphaeriaceae*) and *Biscogniauxia mediterranea* on forest trees in Guilan Province (Iran). *J. Phytopathol.* 2021. V. 169. P. 129–138.
<https://doi.org/10.1111/jph.12967>
- Knudsen H., Vesterholt J. *Funga Nordica*. Agaricoid, boletoid, clavarioid, cyphelloid and gastroid genera. Nordsvamp, Copenhagen, 2012.
- Koukol O., Haňáčková Z., Dvořák M. et al. Unseen, but still present in Czechia: *Hymenoscyphus albidus* detected by real-time PCR, but not by intensive sampling. *Mycol. Progress.* 2016. V. 15. Art. 6.
- Kowalski T., Bilański P. *Hymenoscyphus pusillus*, a new species on leaves of *Fraxinus pennsylvanica* in Poland. *Forest Pathol.* 2019. V. 49 (1). e12481.
- Kowalski T., Holdenrieder O. The Teleomorph of *Chalara fraxinea*, the causal agent of ash dieback. *Forest Pathol.* 2009. Vol. 39 (5). P. 304–308.
- Kumar S., Stecher G., Li M. et al. MEGA X: Molecular Evolutionary Genetics Analysis across computing platforms. *Molecular Biology and Evolution.* 2018. Vol. 35. P. 1547–1549
- Kuprevich V.F., Ulyanishchev V.I. Keys to rust fungi of the USSR. Part 1. Family *Melampsoraceae* and some *Pucciniaceae* species. Nauka i tekhnologiya, Minsk, 1975. (In Russ.).
- Kuzmichev E.P., Sokolova E.S., Kulikova E.G. Common fungal diseases of Russian forests. Washington, D.C., U.S. Dep. of Agriculture, 2001. P. 60.
- Mamaev S.A. Keys to the trees and shrubs of the Urals. Native and alien species. UrB RAS, Ekaterinburg, 2000. (In Russ.).
- Mukhin V.A., Tretyakova A.S., Pryadein et al. Plants and fungi of the National Park “Pripishminsky Bory.” Izdatelstvo Uralskogo universiteta, Ekaterinburg, 2003. (In Russ.).
- Musolin D.L., Selikhovkin A.V., Shabunin D.A. et al. Between ash dieback and emerald ash borer: two Asian invaders in Russia and the future of ash in Europe. *Baltic Forestry.* 2017. V. 23 (1). P. 316–333.
- Naumov N.A. Fungi of the Urals. Notes of UOLE. 1915. V. 35. P. 1–53. (In Russ.).
- Ostry M.E. *Hypoxylon* canker. In: *Infectious forest diseases*. CABI, Wallingford, 2013, pp. 407–419.
- Ostry M.E., Anderson N.A. Genetics and ecology of the *Entoleuca mammata* – *Populus* pathosystem: Implications for aspen improvement and management. *Forest Ecology and Management.* 2009. V. 257 (2). P. 390–400.
<https://doi.org/10.1016/j.foreco.2008.09.053>
- Pacia A., Borowik P., Hsiang T. et al. Ash dieback in forests and rural areas – history and predictions. *Forests.* 2023. V. 14 (11). Art. 2151.
<https://doi.org/10.3390/f14112151>
- POWO, 2025. Plants of the World Online <https://powo.science.kew.org/>. Accessed 29.06.2025.
- Rabitsch W., Nehring S. (Hrsg.) *Naturschutzfachliche Invasivitäts-bewertungen für in Deutschland wild lebende gebietsfremde terrestrische Moose, Flechten und Pilze*. BfN-Skripten 603. Bonn, 2021.
- Ragazzi A., Ginetti B., Moricca S. First Report of *Biscogniauxia mediterranea* on English Ash in Italy. *Disease Notes.* 2012. V. 96 (11). P. 1694.
<https://doi.org/10.1094/pdis-05-12-0442-pdn>
- Ranjana R., Basu M.B. *Biscogniauxia mediterranea* – a wood rotting pathogen. *J. Mycopathol. Research.* 2012. V. 50 (2). P. 337–339
<https://doi.org/10.5555/20133097838>
- Ryvarden L., Melo I. Poroid fungi of Europe. *Synopsis Fungorum.* 2014. V. 31. P. 1–455.
- Schoebel C.N., Zoller S., Rigling D. Detection and genetic characterisation of a novel mycovirus in *Hymenoscyphus fraxineus*, the causal agent of ash dieback. *Infect Genet Evol.* 2014. V. 28. P. 78–86.
- Senwana C., Wanasinghe D.N., Bulgakov T.S. et al. Towards a natural classification of *Dothidotthia* and *Thyrostroma* in *Dothidotthiaceae* (*Pleosporineae*, *Pleosporales*). *Mycosphere.* 2019. V. 10 (1). P. 701–738.
<https://doi.org/10.5943/mycosphere/10/1/15>
- Shiryaev A.G. Changes in mycobiota of Ural-and-Siberian region under global warming and anthropogenic impact. *Bulletin of Ecology, Forest and Landscape Sciences.* 2009. N 9. P. 37–47. (In Russ.).
- Shiryaev A.G., Kotiranta H., Mukhin V.A. et al. Aphyllophoroid fungi of Sverdlovsk Region (Russia). *Biodiversity, distribution, ecology and the IUCN threat categories*. Goschitsky Publ., Ekaterinburg, 2010.
- Shiryaev A.G., Stavishenko I.V. New and rare for Sverdlovsk Region species of basidiomycetes. *Mikologiya i fitopatologiya.* 2011. V. 45 (4). P. 345–349. (In Russ.).
- Shiryaev A.G., Zmitrovich I.V., Shiryaeva O.S. Species richness of *Agaricomycetes* on hedge vines in Ekaterinburg City (Russia). *Mikologiya i fitopatologiya.* 2021. V. 55 (5). P. 340–352.
<https://doi.org/10.31857/S0026364821050093>
- Shiryaev A.G., Zmitrovich I.V., Bulgakov T.S. et al. Global warming favors the development of a rich and heterogeneous mycobiota on alien vines in a boreal city under continental climate. *Forests.* 2022a. V. 13 (2). Art. 323.
<https://doi.org/10.3390/f13020323>
- Shiryaev A.G., Zmitrovich I.V., Shiryaeva O.S. New and rare *Agaricomycetes* species on woody alien plants in Ekaterinburg City (Russia). *Mikologiya i fitopatologiya.* 2022b. V. 56 (5). P. 350–356. (In Russ.)
<https://doi.org/10.31857/S0026364822050105>
- Shiryaev A.G., Zmitrovich I.V., Zhao P. et al. Fungal diversity of native and alien woody leguminous Plants in the Middle Urals. *Contemporary Problems of Ecology.* 2023a. V. 16 (4). P. 403–425.
<https://doi.org/10.1134/S1995425523040091>

- Shiryaev A.G., Kiseleva O.A.* Greenway planning in Ekaterinburg City: Unaccounted phytopathological problems of the urban strategy project. *Contemporary Problems of Ecology*. 2023. V. 16 (4). P. 509–527. <https://doi.org/10.1134/S199542552304008x>
- Shiryaev A.G., Zmitrovich I.V., Senator S.A. et al.* How poor is aphyllorhizoid fungi diversity in the boreal urban greenhouses of Eastern Europe? *J. Fungi*. 2023b. V. 9 (11). Art. 1116. <https://doi.org/10.3390/jof9111116>
- Shiryaev A.G., Bulgakov T.S., Zmitrovich I.V. et al.* New species of fungi for Sverdlovsk region (The Middle Urals, Russia) on alien and aborigine woody plants. *Mikologiya i fitopatologiya*. 2023c. V. 57 (6). P. 417–424. <https://doi.org/10.31857/S0026364823060107>
- Shiryaev A.G., Bulgakov T.S., Shiryaeva O.S. et al.* Further additions to mycobiota of alien and native woody plants in Sverdlovsk region (Russia, Middle Ural). *Mikologiya i fitopatologiya*. 2024. V. 58 (4). P. 294–302. <https://doi.org/10.31857/S0026364824040035>
- Spaulding P.* Foreign Diseases of Forest Trees of the World. U.S.D.A. Agric. Handbook. V. 197. Wanshington, D.C., U.S. Dep. of Agriculture, 1961. 361 p.
- Stepanova-Kartavenko N.T.* Aphyllorhizoid Fungi of the Urals. Sverdlovsk, UFAS USSR, 1967. 295 p. (In Russ.)
- Stepanova N.T., Sirko A.V.* Flora of ascomycetes and imperfect fungi of the Urals. In: *Spore plants of the Urals: Proc. Inst. Ecol. Plant and Animals*. Iss. 70. Sverdlovsk, UFAS USSR, 1970, pp. 3–52. (In Russ.)
- Syuzev P.V.* Materials on the mycological flora of the Perm region. *Bull. Soc. Imp. Nat. Moscou*. 1898. V. 12. P. 320–328. [In Russ.; Latin pt “Enumeratio fungorum, quos in provincia Permensi (in Ural) P. Süsew legit”].
- Syuzev P.V.* Fungal parasites causing diseases to cultural and useful plants in the Perm region. *Materialy po izucheni-yu Permskogo kraja*. 1911. P. 151–158. (In Russ.)
- Teterevnikova-Babayan D.N.* Fungi of the genus *Septoria* in the USSR. Publishing House of the Academy of Sciences of the Armenian SSR, Yerevan, 1987. (In Russ.)
- Ulyanishchev V.I.* Keys to rust fungi of the USSR. Pt 2. *Nauka i Tekhnologiya*, Minsk, 1975. (In Russ.)
- van der Merwe R., Halleen F., van Dyke M. et al.* Occurrence of canker and wood rot pathogens on stone fruit propagation material and nursery trees in the Western Cape in South Africa. *Plant Diseases*. 2021. V. 105 (11). P. 3586–3599. <https://doi.org/10.1094/PDIS-10-20-2124-RE>
- Voglmayr H., Schertler A., Essl F. et al.* Alien and cryptogenic fungi and oomycetes in Austria: an annotated checklist (2nd edition). *Biol. Invasions*. 2023. Vol. 25. P. 27–38.
- White T.J., Bruns T., Lee S. et al.* Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: *M.A. Innis etc. (eds). PCR Protocols: a guide to methods and applications*. Academic Press, N.Y., 1990, pp. 315–322.
- Zhao Y.J., Hosoya T., Baral H.O. et al.* *Hymenoscyphus pseudoalbidus*, the correct name for *Lambertella albidus* reported from Japan. *Mycotaxon*. 2013. V. 122. P. 25–41.
- Zheng H.D., Zhuang W.Y.* *Hymenoscyphus albidoides* sp. nov. and *H. pseudoalbidus* from China. *Mycol. Progress*. 2014. V. 13. P. 625–638. <https://doi.org/10.1007/s11557-013-0945-z>
- Zviagintsev V.B., Demidko D.A., Panteleev S.V. et al.* Distribution of invasive pathogen of ash dieback disease *Hymenoscyphus fraxineus* in European part of Russia. *Izvestia Sankt-Peterburgskoy Lesotekhnicheskoy Akademii*. 2023. V. 244. P. 88–117. (In Russ.). <https://doi.org/10.21266/2079-4304.2023.244.88-117>
- Благовещенская Е.Ю.* (Blagoveshchenskaya) Фитопатогенные микромицеты: учебный определитель. Москва: URSS, 2015. 232 с.
- Булгаков Т.С., Васильев Н.П., Змитрович И.В.* (Bulgakov et al.) Итоги 10-летнего обследования микобиоты пород-интродуцентов дендрария научно-опытной станции „Отрадное“ Ботанического института им. В.Л. Комарова РАН // *Ботаника: история, теория, практика* (к 300-летию основания Ботанического института им. В.Л. Комарова Российской академии наук): Труды международной научной конференции. СПб.: Изд-во СПбГЭТУ „ЛЭТИ“, 2014. С. 31–39.
- Демидова З.А.* (Demidova) Краткий обзор болезней культурных и дикорастущих растений в Уральской области // *Бюллетень УралОблЗУ*. 1925. № 10. С. 9–20.
- Демидова З.А.* (Demidova) О головневых грибах Урала // *Тр. Ин-та экологии раст. и животных*. Вып. 70. 1970. С. 53–60.
- Звягинцев В.Б., Демидко Д.А., Пантелеев С.В. и др.* (Zviagintsev et al.) Распространение инвазивного возбудителя некроза ветвей ясеня аскомицета *Hymenoscyphus fraxineus* в европейской части России // *Изв. Санкт-Петербургской лесотехнической академии*. 2023. Вып. 244. С. 88–117.
- Каратыгин И.В.* (Karatygin) Определитель грибов России. Порядки Taphrinales, Protomycetales, Exobasidiales, Microstromatales. СПб.: Наука, 2002. 135 с.
- Купревич В.Ф., Ульянищев В.И.* (Kuprevich, Ulyanishchev) Определитель ржавчинных грибов СССР. Ч. 1. Сем. Melampsoraceae и некоторые виды Russiniaceae. Минск: Наука и техника, 1975. 336 с.
- Мамаев С.А.* (Mamaev) Определитель деревьев и кустарников Урала. Местные и нтродуцированные виды. Екатеринбург: УрО РАН, 2000. 260 с.
- Мухин В.А., Третьякова А.С., Прядеин Д.В.* Растения и грибы национального парка “Припышминские боры”. Екатеринбург: Изд-во Уральского ун-та. 2003. 205 с.
- Наумов Н.А.* (Naumov) Грибы Урала // *Зап. УОЛЕ*. 1915. Т. 35. С. 1–53.
- Степанова-Картавенко Н.Т.* (Stepanova) Афиллофоровые грибы Урала. Свердловск: УФАН СССР, 1967. 295 с.
- Степанова Н.Т., Сирко А.В.* (Stepanova, Sirko) К флоре сумчатых и несовершенных грибов Урала // *Споровые растения Урала: Тр. инст. экол. раст. и жив.* Вып. 70. Свердловск: УФАН СССР, 1970. С. 3–52.
- Сюзев П.В.* (Syuzev) Материалы к микологической флоре Пермской губернии // *Bulletin de la Société impériale des naturalistes de Moscou*. 1898. Т. 12. С. 320–328.
- Сюзев П.В.* (Syuzev) Грибные паразиты, причиняющие болезнь культурным и полезным растениям в Пермской губернии // *Мат-лы по изуч. Пермского края*. 1911. Вып. 4. С. 151–158.

- Тетеревникова-Бабаян Д.Н. (Teterevnikova-Babayan) Грибы рода *Septoria* в СССР. Ереван: АН АрмССР, 1987. 479 с.
- Ульянищев В.И. (Ulyanishchev) Определитель ржавчинных грибов СССР. Ч. 2. Л.: Наука, 1978. 384 с.
- Ширяев А.Г. (Shiryaev) Изменения микобиоты Урало-Сибирского региона в условиях глобального потепления и антропогенного воздействия // Вест. экологии, лесоведения и ландшафтоведения. 2009. № 9. С. 37–47.
- Ширяев А.Г., Ставишенко И.В. (Shiryaev, Stavishenko) Дополнение к списку видов базидиальных грибов Свердловской области // Микология и фитопатология. 2011. Т. 45. Вып. 4. С. 345–349.
- Ширяев А.Г., Змитрович И.В., Ширяева О.С. (Shiryaev et al.) Новые и редкие виды агарикомицетов на древесных интродуцентах в г. Екатеринбурге (Россия) // Микология и фитопатология. 2022. Т. 56. Вып. 5. С. 350–356.

Новые и редкие виды грибов, развивающиеся на интродуцированных древесно-кустарниковых растениях в Свердловской области (Средний Урал, Россия)

О. С. Ширяева^{a, #}, Т. А. Сурина^{b, ##}, И. В. Змитрович^{c, ###}, А. С. Будимиров^{a, ####},
Т. С. Булгаков^{d, #####}, Л. В. Зайцева^{b, #####}, А. Г. Ширяев^{a, #####}

^a Институт экологии растений и животных УрО РАН, Екатеринбург, Россия

^b Всероссийский центр карантина растений, Быково, Россия

^c Ботанический институт им. В.Л. Комарова РАН, Санкт-Петербург, Россия

^d Федеральный исследовательский центр “Субтропический научный центр РАН”, Сочи, Россия

[#]e-mail: olga.s.shiryaeva@gmail.com

^{##}e-mail: t.a.surina@yandex.ru

^{###}e-mail: iv_zmitrovich@mail.ru

^{####}e-mail: budimirov.alex@gmail.com

^{#####}e-mail: ascomycologist@yandex.ru

^{#####}e-mail: lidiya.zaitzeva26@yandex.ru

^{#####}e-mail: anton.g.shiryaev@gmail.com

Впервые для Свердловской обл. приводятся сведения о находках 28 видов фитопатогенных и сапротрофных высших грибов (*Ascomycota* и *Basidiomycota*), найденных на интродуцированных видах деревьев и кустарников. Образцы еще 14 редких видов собраны в регионе во второй — четвертый раз. Четыре вида — *Cheirospora botryospora*, *Homortomyces tamaricis*, *Lophomerum rhododendri* и *Pyrenopeziza lonicerae* — обнаружены в России впервые. Грибы собраны на 34 видах древесных и кустарниковых растений, включая 27 видов, развивающихся в открытом грунте, и семь в оранжереях. Приводятся сведения о местонахождениях опасных фитопатогенных грибов: *Biscogniauxia mediterranea*, *Entoleuca mammata*, а также *Hymenoscyphus fraxineus*, для которого выявлено новое местонахождение. Анализ последовательности ITS1–5.8S–ITS2, полученной из собранных в ходе текущего исследования апотециев *H. fraxineus*, показал заметные отличия от европейских популяций этого вида и продемонстрировал сходство с восточноазиатским вариантом последовательности. Таким образом, вновь найденные местные варианты *H. fraxineus* в Свердловской обл. могут иметь не европейское, а восточноазиатское происхождение.

Ключевые слова: биоразнообразие, инвазия, патогены растений, распространение грибов, чужеродные виды, *Ascomycota*, *Basidiomycota*